PYRROLIZIDINE ALKALOIDS FROM SENECIO CACALIASTER*

ERHARD ROEDER, HELMUT WIEDENFELD and RUTH BRITZ-KIRSTGEN

Pharmazeutisches Institut der Universitat Bonn, An der Immenburg 4, D-5300 Bonn 1, West Germany

(Revised received 30 January 1984)

Key Word Index—Senecio cacaliaster; Compositae; pyrrolizidine alkaloids; O⁷-senecioylretronecine; O⁷-senecioyl-sarracinyl-retronecine; sencalenine; bulgarsenine.

Abstract—Four pyrrolizidine alkaloids have been isolated from *Senecio cacaliaster* and their structures analysed by spectroscopic methods (IR, mass, ¹H, ¹³C NMR). One of them is new and the name sencalenine (3) is proposed. Alkaloids O⁷-senecioylretronecine (1) and 7-senecioyl-9-sarracinylretronecine (2) have recently been identified elsewhere. The fourth is bulgarsenine (4) which was isolated from a *Senecio* species before.

INTRODUCTION

Senecio cacaliaster is widely distributed in Europe. On account of the fact that this plant possesses no ligules, Linné adjoined it to the genus Cacalia and named it S. cacaliaster. However, in spite of the missing ligules this plant does resemble S. nemorensis, especially S. nemorensis subsp. fuchsii which may be implied from its outward shape. Both of them belong to the Betulo adenostyletea [1]. Ecologically S. cacaliaster is closely related to S. nemorensis subsp. fuchsii, from which the pyrrolizidine alkaloids (PA) fuchsisenecionine and senecionine could be isolated [2, 3]. S. lamottei is a hybrid of these two species. Therefore, a common precursor of S. cacaliaster and S. nemorensis has to be considered. We investigated S. cacaliaster with regard to its alkaloids in order to prove this relationship. We detected four major alkaloids by TLC, which we then separated by droplet counter current chromatography (DCCC).

RESULTS AND DISCUSSION

Methanolic extraction of plant material was followed by purification of the residue as already described [3]. From the resulting residue four PAs have been isolated by DCCC. The mass spectra gave the formulae C₁₃H₁₉NO₃ for 1, $C_{18}H_{25}NO_5$ for 2 and 3 and $C_{18}H_{27}NO_5$ for 4. The fragmentation of the latter alkaloid is identical to that of bulgarsenine [4]. A typical fragmentation of 1-3 between m/z 136 and m/z 80 is characteristic for retronecine or its isomeric form. In addition, the fragmentations show that PAs 2 and 3 are isomeric diesters, whereas PA 1 must be a retronecine monoester. The mass spectrum also gives information about the esterification of PA 1 at C-7 with a 5-C-linked acid. The alkaloids 2 and 3, having similar 5-Clinked acids, are esterified at both alcoholic positions. Likewise, the IR data show for alkaloid 1 one ester bond, whereas in 2, 3 and 4 two such bonds were found. Further information about the structures is given by NMR analysis (Tables 1 and 2).

The NMR data for 1 are reported for the first time; in the case of alkaloid 2 our data differ from those described recently [5] by interchange of the 13 C-values for carbons $^{1/12/16}$ and $^{3/9}$ and $^{13/14}$. Likewise, there is some difference in the 1 H NMR data especially in the values for the C-5 H₂. All values were verified by decoupling experiments and by evaluation of coupled and noise-decoupled spectra. The value for C-6 H₂ is higher by 2 ppm for all three alkaloids. This also demonstrates that retronecine is the necic base [6, 7]. The NMR data indicate the structure of an 07 -senecioic-acid-retronecine-ester for PA 1. This is clear from the 1 H peaks at $^{\delta}$ 1.89 and 2.16 for two methyls and the corresponding

3

^{*}Dedicated to Professor Maximilian Steiner on the occasion of his 80th birthday.

1762 E. ROEDER et al.

Table 1. ¹H NMR data of 1, 2 and 3 (CDCl₃; TMS)

		· ·	•
	1	2	3
C ₁₃ -H ₃	1.89, d, 3H	1.87, d, 3H	2.04, d, 3H
	J = 1	J = 1	
C14-H3	2.16, d, 3H	2.05, d, 3H	_
	J = 1	J = 1	
C14-H2	_	_	3.62, m, 2H
C ₁₄ -OH	_	_	4.73, m, OH
C ₆ -H ₂	2.06, m, 2H	2.67, q, 2H	2.33, m, 2H
• -		J=9	
C ₅ -H _A	2.71, m, 1H	3.31, m, 1H	2.29, m, 1H
C ₃ -H _A	3.28, m, 1H	3.49, m, 1H	3.1, m, 1H
C ₅ -H _B	3.38, m, 1H	3.67, m, 1H	3.6, m, 1H
C ₃ -H _B	3.89, m, 1H	3.93, m, 1H	3.75, m, 1H
C_{18} - H_3	_	2.02, d, 3H	1.97, 2q, 3H
		J = 7.5	J = 7; 1.5
C ₉ -H ₂	4.1, d, 2H	4.78, m, 2H	4.71, m, 2H
C ₁₉ -H ₃			1.87, q, 3H
			J = 1.5
C ₁₉ -H ₂		4.2, s, 2H	
C ₁₉ -OH	_	4.39, m, OH	_
C ₈ -H	4.7, m, 1H	4.51, m, 1H	4.44, m, 1H
C ₉ -OH	4.57, m, OH	-	
C ₇ -H	5.45, m, 1H	5.4, m, 1H	5.62, m, 1H
C ₁₁ -H	5.6, m, 1H	5.6, m, 1H	5.97, m, 1H
C ₂ -H	5.68, m, 1H	5.78, m, 1H	5.78, m, 1H
C ₁₇ -H		6.38, q, 1H	6.16, 2q, 1H
		J=7.5	J = 7

 δ values in ppm; J in Hz.

Table 2. 13NMR data of 1, 2 and 3 (CDCl₃; TMS)

Carbon No.	1	2	3
18	_	15.8	15.9
13	20.46	20.3	20.6
14	27.58	27.5	66.8
6	34.31	34.4	34.2
5	53.47	53.7	54.0
9	59.43	60.8	60.5
3	61.8	62.3	62.1
19		64.7	16.0
7	72.44	72.9	71.8
8	76.32	75.7	76.4
11	115.0	115.8	123.8
2	121.8	127.0	126.8
1	138.79	131.8	140.2
12	138.75	133.9	133.6
17		141.0	128.1
16	_	158.0	160.8
15	_	165.6	164.9
10	168.0	166.6	167.18

^{*} δ values in ppm.

olefinic proton at 5.6. The structure of 1 is also verified from the corresponding ¹³C data. The NMR spectra of PA 2 show partially the same peaks as the NMR data of 1; thus, 1 is found as a partial structure in 2. Furthermore, in 2 the esterified acid at C-9 must be a hydroxyangelic acid.

This is confirmed by a quartet at $\delta 2.02$ with a coupling constant of 7.5 Hz for C-18 H₃ and the olefinic proton at 6.38 with the same coupling constant. The hydroxyl group must be located at position C-19 on account of the value δ 4.2 for the methylene group. This structure is also established by the following 13 C data: δ 64.7 for C-19 and 15.8 for C-18. The data for compound 3 show that it is an isomer of 2. Instead of peaks for the methyl group at C-14 those of a hydroxymethyl group at δ 4.73 (¹H) and 66.8 (13C) are found. In comparison with 2, peaks for an angelic acid are present in 3 for the acid at C-9: C-18 H₃: $\delta 1.97 (J = 7; 1.5 \text{ Hz}) = {}^{1}\text{H} \text{ and } 15.9 = {}^{13}\text{C NMR}; \text{C-}19$ H_3 : $\delta 1.87$ (J = 1.5 Hz) = ¹H and $16.0 = ^{13}$ C NMR. The esterification of 2 and 3 as shown in the formulae is proved by mass spectrometry. The fragmentation pattern of 2 shows an intense ion at m/z 220. This ion is formed after ester cleavage at C-9 building a necine, esterified at C-7 with a 3-methyl-2-butenoic acid. Furthermore, ion m/z219 shows a McLafferty rearrangement and a loss of 100 mu (m/z 119). The ion m/z 237 also proves the structure 2. It is formed after rearrangement of the ester grouping at C-9 and the loss of C₅H₆O₂ indicating the hydroxymethyl group position at C-1. In the fragmentation pattern of 3 the ion m/z 219 is formed after McLafferty rearrangement and the loss of 116 mu. Only alkaloids which possess an open diester form show such rearrangements at C-7. Therefore, in 3 there must be an acid of MW 116 esterified at C-7. Furthermore, an ester cleavage is found at C-9 forming the ion m/z 236. This esterification at C-7 and C-9 in 3 is established by the fact that the ion m/z 220 is missing. For the fourth alkaloid (= bulgarsenine) we found similar data as reported earlier [4, 8]. For the new alkaloid 3 we propose the name sencalenine.

EXPERIMENTAL

S. cacaliaster (Lam.) was collected in alpine Austria (Highlands of Grossglockner). The dried and pulverized drug was extracted and purified as mentioned earlier [3]. The resulting yellow residue was separated by DCCC using toluene–MeOH– $\rm H_2O-CHCl_3$ in the ascending mode yielding the four PAs [9]. Content of PAs (GC; dry wt): 1, 0.017%; 2, 0.005%; 3, 0.005%; 4, 0.21%. GC conditions: glass column: 1.8 m, 2 mm; 4% SE 30 on Varaport 30; prog: 80–240° in four steps at 9°/min: 2 × 2°/min; 1 × 3°/min; 1 × 4°/min. Inj.: 200°. Det.: 260°; carrier gas (N₂): 40 ml/min. $R_{\rm Sr}$ values (min): 1, 1.53; 2, 21.8; 3, 17.6; 4, 20.15.

 O^7 -Senecioylretronecine (1). Oily; $[\alpha]_D^{20} = -2$ (EtOH). IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3385 (OH), 3010 (C=C), 1730 (satd ester), 1705 (unsatd ester), 1655 (C=C). CIMS (CHCl₃), 70 eV, m/z (% rel. int.): 237.137 [M]⁺ (10.29), 219 [M-H₂O]⁺ (1.47), 154 [M-C₅H₇O]⁺ (11.05), 137 [M-C₅H₈O₂]⁺ (38.64), 136 [219-C₅H₇O]⁺ (30.76), 124 [154-CH₂O]⁺ (34.31), 111 [137-C₂H₂]⁺ (47.09), 106 [137-CH₂-OH]⁺ (48.94), 94 [111-OH]⁺ (24.77), 80 [111-CH₂-OH]⁺ (100.0).

7-Senecioyl-9-sarracinylretronecine (2). Oily; $[\alpha]_{D}^{20} = +4$ (EtOH). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3390 (OH), 3010 (C=C), 1720 (satd ester) 1705 (unsatd ester), 1645 (C=C). CIMS (CHCl₃), 70 eV, m/z (rel. int.): 335.173 [M]⁺ (9.06) 237 [M-C₅H₆O₂]⁺ (42.32), 221 [M-C₅H₆O₃]⁺ (7.08), 220 [M-C₅H₇O₃]⁺ (40.85), 219 [M-C₅H₈O₃]⁺ (8.79), 138 [221-C₅H₇O]⁺ (12.81), 137 [220-C₅H₇O]⁺ (11.96), 136 [219-C₅H₇O]⁺ (100.0), 121 [221-C₅H₈O₂]⁺ (17.59), 120 [220-C₅H₈O₂]⁺ (37.08), 119 [219-C₅H₈O₂]⁺ (28.93), 106 [137-CH₃O]⁺ (8.67), 95 [121-C₂H₂]⁺ (7.83), 94 [120-C₂H₂]⁺ (46.57), 93 [119-26]⁺ (67.31).

Sencalenine (3). Oily; $[\alpha]_D^{20} = -8$ (EtOH). IR v_B^{Mpx} cm⁻¹: 3380 (OH), 3060 (C=C), 1730 (satd ester), 1710 (unsatd ester), 1660 (C=C). CIMS (CHCl₃) 70 eV, m/z (rel. int.): 335.172 [M] $^+$ (13.50), 252 $[M-C_5H_7O_2]^+$ (22.04), 236 $[M-C_5H_7O_2]^+$ (28.46), 235 $[M-C_5H_8O_2]^+$ (15.16), 219 $[M-C_5H_8O_3]^+$ (12.32), 154 $[252-C_5H_6O_2]^+$ (20.18), 137 $[219-C_5H_6O]^+$ (11.88), 136 $[235-C_5H_7O_2]^+$ (100.0), 122 $[237-C_5H_7O_3]^+$ (28.10), 120 $[219-C_5H_7O_2]^+$ (52.06), 119 $[235-C_5H_8O_3]^+$ (31.13), 106 $[137-CH_2-OH]^+$ (9.40), 94 $[120-C_2H_2]^+$ (60.88), 93 $[119-C_2H_2]^+$ (79.55), 80 $[122-C_3H_6]^+$ (18.27).

Acknowledgement—We thank the Deutsche Forschungsgemeinschaft for financial support.

REFERENCES

- Ellenberg, H. (1978) Vegetation Mitteleuropas mit den Alpen, p. 571. Verlag Eugen Ulmer, Stuttgart.
- 2. Roeder, E. and Wiedenfeld, H. (1977) Phytochemistry 16, 1462.
- 3. Wiedenfeld, H. and Roeder, E. (1979) Phytochemistry 18, 1083.
- Nguyen Thi Nghia, Sedmera, P., Klasek, A., Boeva, A., Drjanovska, L., Dolejs, L. and Santavy, F. (1976) Coll. Czech. Chem. Commun. 41, 2952.
- 5. Rüeger, H. and Benn, M. H. (1983) Can. J. Chem. 61, 2526.
- Bull, L. B., Culvenor, C. C. J. and Dick, A. T. (1968) The Pyrrolizidine Alkaloids, p. 40. North Holland, Amsterdam.
- Culvenor, C. C. J. and Woods, W. G. (1965) Aust. J. Chem. 18, 1625.
- Roeder, E., Wiedenfeld, H. and Frisse, M. (1980) Phytochemistry 19, 1275.
- 9. Otsuka, H., Ogihara, Y. and Shibata, S. (1974) Phytochemistry 13, 2016.